



**Cite this article:** Wu J, Jiao H, Simmons NB, Lu Q, Zhao H. 2018 Testing the sensory trade-off hypothesis in New World bats. *Proc. R. Soc. B* **285**: 20181523. <http://dx.doi.org/10.1098/rspb.2018.1523>

Received: 5 July 2018

Accepted: 4 August 2018

**Subject Category:**

Evolution

**Subject Areas:**

evolution, ecology, genetics

**Keywords:**

sensory trade-off, pseudogene, opsin, echolocation, vampire bat

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Electronic supplementary material is available online at <https://dx.doi.org/10.6084/m9.figshare.c.4196597>.

# Testing the sensory trade-off hypothesis in New World bats

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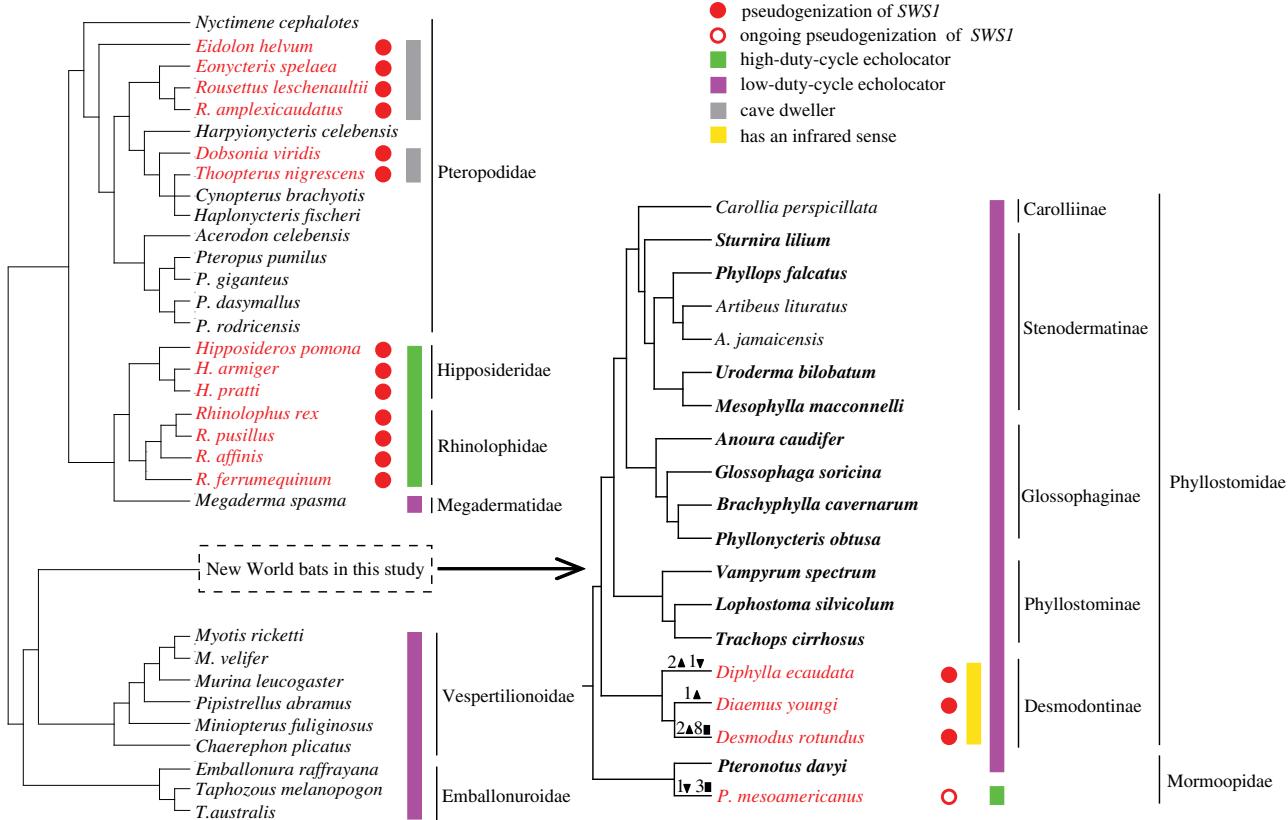
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Detection of evolutionary shifts in sensory systems is challenging. By adopting a molecular approach, our earlier study proposed a sensory trade-off hypothesis between a loss of colour vision and an origin of high-duty-cycle (HDC) echolocation in Old World bats. Here, we test the hypothesis in New World bats, which include HDC echolocators that are distantly related to Old World HDC echolocators, as well as vampire bats, which have an infrared sensory system apparently unique among bats. Through sequencing the short-wavelength opsin gene (*SWS1*) in 16 species (29 individuals) of New World bats, we identified a novel *SWS1* polymorphism in an HDC echolocator: one allele is pseudogenized but the other is intact, while both alleles are either intact or pseudogenized in other individuals. Strikingly, both alleles were found to be pseudogenized in all three vampire bats. Since pseudogenization, transcriptional or translational changes could separately result in functional loss of a gene, a pseudogenized *SWS1* indicates a loss of dichromatic colour vision in bats. Thus, the same sensory trade-off appears to have repeatedly occurred in the two divergent lineages of HDC echolocators, and colour vision may have also been traded off against the infrared sense in vampire bats.

## 1. Introduction

Sensing the ever-changing environment is a fundamental task that all organisms must accomplish to survive in nature. Depending on the species, animals may rely on distinct sets of sensory modalities, with some species lacking one or more basic senses while others develop novel senses [1–3]. Detection of evolutionary shifts in sensory systems is challenging in animals, possibly due to difficulties in observing and measuring sensory functions [4]. By adopting a molecular approach, one group of animals was found to show losses of colour vision—the bats (order Chiroptera) [5], which represent approximately 20% of all living mammal species [6]. Specifically, members of the suborder Yangochiroptera within Chiroptera possess dichromatic colour vision, while some members of the other suborder, Yinpterochiroptera, have lost the ability to distinguish colours [5]. Within Yinpterochiroptera, independent losses of colour vision in several species of the Old World fruit bats (Pteropodidae) may be associated with a behavioural innovation—the choice of caves as roost sites in a clade that otherwise roosts in trees [5]. Loss of colour vision has also occurred independently in another lineage composed of two sister clades of Old World bats, Rhinolophidae and Hipposideridae [5]. This loss apparently coincided with the origin of a novel sensory modality, high-duty-cycle (HDC) echolocation, a sensory specialization that facilitates hunting prey in dense vegetation by separating frequency of emitted call pulses and returning echo [5]. Most echolocating bats use low-duty-cycle (LDC) echolocation and a separate pulse and echo in time, a system that is widely thought to be a primitive in bats [7–9]. As far as is known, the majority of LDC bats also have dichromatic colour vision [5]. As a result of this pattern, a sensory trade-off between the loss of colour vision and the origin of the HDC echolocation in bats has been hypothesized [5]. Under this hypothesis, convergent losses of colour



**Figure 1.** The species tree depicting the evolution of *SWS1* in bats. The tree topology follows previous studies [17–19]. Species shown in the left panel were derived from Zhao *et al.* [5], while the species detailed in the right panel are the New World bats used in this study; In the New World bats, the 16 species newly sequenced are shown in bold font, whereas the three species previously sequenced are shown in regular font. Numbers of ORF-disrupting insertions (filled inverted triangle), deletions (filled triangle) and premature stops (filled square) are shown on each branch. Species indicated by green bars are high-duty-cycle echolocators, while species indicated with purple bars are low-duty-cycle echolocators; species indicated with the yellow bar are vampire bats that possess infrared sense. Species shown in red possess *SWS1* pseudogenes and thus lack typical dichromatic colour vision, and these species were further divided into three categories indicated by three colour bars: green (high-duty-cycle echolocators), yellow (vampire bats) and grey (cave-roosting Old World fruit bats).

vision with gains of other sensory modalities might be expected to have independently occurred in distantly related lineages. However, other authors have argued that the coincidence of HDC echolocation and loss of colour vision was just a mere coincidence [10]. Addressing these competing hypotheses requires additional data and analyses.

To test the sensory trade-off hypothesis, we turn to the New World bats. These species include the moustached bat (*Pteronotus parnellii*) and vampire bats. The former has independently evolved HDC echolocation that is similar to the Old World lineages (hipposiderids and rhinolophids) [7,8], while the latter have evolved a unique sensory modality among bats—the infrared sense [2,11]. In mammals, the cone opsins are typically divided into two categories: middle/long wavelength sensitive (M/LWS) and short wavelength sensitive (SWS) [12]. Most mammals have both the two categories of opsin and thus possess dichromatic colour vision, whereas a small proportion of mammals retain only one category of opsin and thus are monochromatic (a condition also known as colour blindness) [10]. Two types of SWS opsin genes are present in mammals: monotremes have *SWS2*, whereas marsupials and eutherians possess *SWS1* [13,14]. Several studies in mammals have demonstrated conservation of M/LWS but extraordinary divergence of *SWS1* across taxa [5,15,16], we thus focused on the evolution of *SWS1* in this work. Although pseudogenization of coding sequence, transcriptional or translational changes could separately result in functional loss of a gene, we here predict the loss of dichromatic colour vision in

New World bats by identifying *SWS1* pseudogenes (figure 1; electronic supplementary material, table S1). There is no causal link between pseudogenization and functional loss, as the latter may have preceded the former at transcriptional or translational stage. Thus, pseudogenization of *SWS1* can predict a loss of dichromatic colour vision, while an intact *SWS1* cannot predict intact colour vision in bats. Through sequencing the short-wavelength opsin gene (*SWS1*) in 16 species (29 individuals) of New World bats (figure 1; electronic supplementary material, table S1), we inferred losses of dichromatic colour vision to test (i) whether there has been a sensory trade-off between vision and echolocation in *Pteronotus* species similar to that seen in the distantly-related hipposiderid and rhinolophid lineages, and (ii) whether there has been a sensory trade-off between vision and the infrared sense in vampire bats.

## 2. Material and methods

### (a) Taxa and DNA sequencing

Tissues of 16 New World bats (29 individuals) examined in this study were provided by the American Museum of Natural History, and information about these samples is shown in electronic supplementary material, table S1. Our samples contained the HDC echolocator (*Pteronotus mesoamericanus*) and its congeneric species (*Pteronotus davyi*) that uses LDC echolocation, and all three extant species of vampire bats (subfamily Desmodontinae

(a)	human	Exon2 (427)	TTCCGC- <b>TT</b> CAGCTCCAAGCATGCACGTACGGTGGTCCTGGC	
	<i>Pdav</i>		TTCCGC- <b>TT</b> CAGCTCCAAGCACGCCTGATGGTAGTCCTGGC	
	<i>Pmes</i> (intact allele)		TTCCGC- <b>AT</b> CAGCTCCAAGCACGCACGGATGGTAGTCCTGGC	
	<i>Pmes</i> (null allele)		TTCCGC <b>CAT</b> CAGCTCCAAGCACGCACGGATGGTAGTCCTGGC	(insertion: +1 frameshift)
(b)	human	Exon1 (118)	TTCATGGGCACTGTCTCCTTATAGGGTTCCC <b>ACT</b> CAATGCC	
	<i>Drot</i>		TTCATGGGCTTGTCTTGCAGGGAT <b>CC</b> -CTCAATGCC	(deletion: -1 frameshift)
(c)	human	Exon4 (835)	GGGCTGGACTTACGGCTTGTACCAT <b>T</b> CCTTCATTCTCTCC	
	<i>Dyou</i>		GGGCTGGACTTCCGGCTGGTAC <b>AT</b> -CCTGCCTTCTCTCC	(deletion: -1 frameshift)
(d)	human	Exon1 (214)	ATTCTGGTCAACGTGTCTTC- <b>GG</b> AGGGCTTCCTCTGCAT	
	<i>Deca</i>		ATTTGGTCAATGTGTCCCTG <b>A</b> GGGGCTTCCTTTGCAT	(insertion: +1 frameshift)

**Figure 2.** Pseudogenizations of *SWS1* in four species of New World bats. Codons containing the first ORF-disrupting mutations are boxed. Dashes indicate alignment gaps and numbers in parentheses indicate the nucleotide positions following human *SWS1*. The abbreviations of *Pdav*, *Pmes*, *Drot*, *Dyou* and *Deca* represent *Pteronotus davyi*, *Pteronotus mesoamericanus*, *Desmodus rotundus*, *Diaemus youngi* and *Diphylla ecaudata*, respectively. (Online version in colour.)

within Phyllostomidae) and their related species (electronic supplementary material, table S1). Of note, *Pteronotus parnellii* was previously thought to be the only bat species in the New World that uses HDC echolocation [7]. However, recent evidence indicates that *P. parnellii* as traditionally recognized is actually a cryptic species complex comprising at least 9 species [20,21]. *Pteronotus mesoamericanus*, previously considered a subspecies of *P. parnellii*, has been elevated to a full species [20,21]. Although most species ( $n = 13$ ) in our samples have only one individual, three species have either five (*Desmodus rotundus* and *P. davyi*) or six individuals (*P. mesoamericanus*) (electronic supplementary material, table S1). Short-wavelength opsin gene (*SWS1*) spanning from exon 1 to exon 4 (about 2.2 kb in length) was amplified and directly sequenced in both directions. In addition, we amplified and sequenced the *M/LWS* opsin gene spanning from exon 1 to exon 6 (about 14 kb in length) in the same five individuals of *Desmodus rotundus* as those examined *SWS1*. All primer sequences used in this study are provided in electronic supplementary material, table S2.

### (b) Phylogenetic and evolutionary analysis

Deduced protein sequences were aligned by the BioEdit program [22] with the ClustalW multiple alignment option; the nucleotide sequence alignments were generated according to the protein sequence alignments and carefully checked by eye. A species tree of the bats in our study was derived from previous studies [17–19]. Phylogenetic reconstruction for the *SWS1* dataset was conducted using a Bayesian approach implemented in MrBayes (version 3.2.6) [23] with the best-fitting model of sequence evolution predicted by MODELTEST (version 2.3) [24]. PHYLML (version 3.2) was used to reconstruct the maximum-likelihood (ML) tree of the *SWS1* dataset under the HKY+I model that was inferred from jMODELTEST (version 2.1.10) [25] with 100 bootstrap replicates. Both Kishino–Hasegawa and Shimodaria–Hasegawa tests were performed to test the difference between the ML/Bayesian trees and the established species tree using Tree-puzzle [26].

To visualize the average rates of nonsynonymous ( $d_N$ ) and synonymous ( $d_S$ ) substitutions per site for sequences with open reading frame (ORF)-disrupting indels, the Nei and Gojobori method [27] implemented in SWAAP 1.0.2 [28] was applied.

We estimated the variation in  $\omega$  (the ratio of nonsynonymous to synonymous substitution rates) along the phylogeny and examined differential selective pressures using PAML [29]. We inferred ancestral sequences with a combination of the likelihood-based method in PAML [29,30] and the maximum-parsimony approach [31]. We calculated the selection intensity parameter ( $k$ ) and detected relaxed selection using RELAX [32] implemented in HyPhy [33]. See also electronic supplementary material.

### (c) Pseudogene dating analysis

We investigated when the functional constraint on *SWS1* became relaxed in each pseudogenized sequence. We assumed that the functional relaxation on *SWS1* started at  $t$  million years ago, and the pseudogenization dates were estimated based on two different methods described in Meredith *et al.* [34] and Zhao *et al.* [35], respectively. See also electronic supplementary material.

## 3. Results

### (a) Pseudogenization of *SWS1* in the New World high-duty-cycle echolocator

Despite the conservation of *SWS1* in most New World bats (see also electronic supplementary material), we found frame-shifting mutations in the New World HDC echolocator (*P. mesoamericanus*). We first amplified the *SWS1* gene from one individual of this species, and the PCR products were cloned and sequenced. We found that sequences from several clones possess intact ORFs (electronic supplementary material, figure S1a), while one 1 bp insertion was present in the sequences of other clones (electronic supplementary material, figure S1b). The 1 bp insertion in the heterozygous state was further confirmed by direct sequencing in the same individual (electronic supplementary material, figure S1c). The 1 bp insertion created one premature stop codon at the beginning of the fifth transmembrane domain (figure 2; electronic supplementary material, figure S2), and the resulting *SWS1* opsin would lack the final three transmembrane

domains and thus be nonfunctional. Sliding window analysis also suggested relaxation of functional constraint, because a sudden increase of substitution rates was observed right after the occurrence of the 1 bp insertion (electronic supplementary material, figure S3). We next examined variation in the *SWS1* gene using five additional individuals of *P. mesoamericanus*. A total of six individuals were sequenced: four were found to have the 1 bp insertion in the heterozygous state, one has the 1 bp insertion in the homozygous state, and one was intact for both alleles. Thus, the percentage of this null allele (with the presence of the 1 bp insertion) in six individuals was 50%, indicating that the insertion has not been fixed in populations and the *SWS1* gene in *P. mesoamericanus* is probably under ongoing pseudogenization (electronic supplementary material, figures S2 and S3). For comparison, we additionally sequenced a congeneric species *P. davyi* that is closely related to *P. mesoamericanus* but uses the LDC echolocation (figure 1). Five individuals of *P. davyi* were examined and we found that all of these bats possess an intact *SWS1* gene and none contained any frameshifting or nonsense mutations (figure 2; electronic supplementary material, figure S2). These results suggest that the null allele is specific to the New World HDC echolocator (*P. mesoamericanus*).

### (b) Pseudogenization of *SWS1* in all three species of vampire bats

Strikingly, all three extant vampire bats were found to possess ORFs of *SWS1* that are disrupted by indels (insertions and/or deletions) (figure 2; electronic supplementary material, figure S4). Specifically, the common vampire bat (*Desmodus rotundus*) has two 1 bp deletions, one 12 bp deletion and one 3 bp deletion, which together resulted in eight premature stop codons (electronic supplementary material, figure S4). The first premature stop codon is located at the end of the first transmembrane domain, which would lead to the loss of the remaining six transmembrane domains of its protein. Despite multiple attempts, we were only able to sequence a short fragment (298 bp) of the *SWS1* gene in the white-winged vampire bat (*Diaemus youngi*) due to failed amplifications. The sequenced region contains one 1 bp deletion that would result in the disruption of the last transmembrane domain (figure 2; electronic supplementary material, figure S4). The hairy-legged vampire bat (*Diphylla ecaudata*) has one 1 bp insertion, one 1 bp and one 2 bp deletion, causing frameshifts starting from the second transmembrane domain (figure 2; electronic supplementary material, figure S4). Although premature stop codons were not observed, both vampire bats (*Diphylla ecaudata* and *Diaemus youngi*) are likely to have a non-functional *SWS1* gene, a conclusion that is also supported by our sliding window analysis (electronic supplementary material, figure S3). Indeed, an elevated  $\omega$  ratio was observed at or downstream of the frameshifting mutations (electronic supplementary material, figure S3), suggesting the strong impact of the ORF-disrupting indels on the gene functionality, even where premature stop codons were not recorded (electronic supplementary material, figure S3b,c). Sliding window analysis also recorded occurrences of ORF-disrupting indels after the  $\omega$  ratio has increased (electronic supplementary material, figure S3c), suggesting the genetic divergence is at least partially shaped by relaxed selection. Although the sequenced region of *SWS1* in *Diaemus youngi* is short, an apparent high  $\omega$  ratio was observed ahead of the location of the single

frameshifting indel (electronic supplementary material, figure S3c), suggesting that the *SWS1* gene in this lineage is under strong relaxed selection and likely nonfunctional. In addition, the relaxed selection on *SWS1* in all three vampire bats was further confirmed by the results recovered from the programs PAML and RELAX (electronic supplementary material, table S3). Notably, no shared mutations were found among any pair of the three species, despite the fact that they all possess frame-shifting mutations and share a recent common ancestry [18]. We were unable to examine multiple individuals of *Diphylla ecaudata* and *Diaemus youngi*, but we examined five individuals of *Desmodus rotundus* and found them to each have the same indels in both alleles. Thus, we infer that all ORF-disrupting mutations observed in *Desmodus rotundus* are likely fixed in its population.

Given the *SWS1* gene is pseudogenized in all three vampire bat species, we attempted to determine when the relaxation of functional constraint on *SWS1* started in this clade. To this end, we estimated the selective pressures on *SWS1* among vampire bats and tested whether there is a signal of relaxation in their most recent common ancestor. The ancestral sequence for vampire bats was inferred with the combination of Bayesian and parsimony approaches [36]. Under the assumption of the same  $\omega$  for all branches,  $\omega_0$  was assumed to be 0.16 (model C, electronic supplementary material, table S3), indicative of purifying selection acting on *SWS1* across all studied bats. We compared model C with model D (assuming the ancestral branch of vampire bats has  $\omega_1$  while other non-vampire bats branches have  $\omega_0$ ), and found that  $\omega_1$  is not significantly different from  $\omega_0$  ( $p = 1$ , electronic supplementary material, table S3), suggesting similar levels of selective pressure on *SWS1* between the common ancestor of vampire bats and other bats. Similarly, the results recovered from RELAX program also revealed that selective pressure on *SWS1* is not relaxed along the common ancestor of vampire bats compared with other lineages ( $k = 1.128$ ;  $p = 1$ , electronic supplementary material, table S3). Together, these findings suggest that *SWS1* was under purifying selection on the ancestral branch leading to vampire bats, and that the relaxation of functional constraint leading to pseudogenization of *SWS1* in the three extant vampire lineages may have occurred recently.

### (c) Dating the *SWS1* pseudogenization events in the New World bats

To date the pseudogenization events in New World bats, we estimated when the functional constraint became relaxed for each of the four pseudogenized *SWS1* sequences, including three vampire bats (*Desmodus rotundus*, *Diaemus youngi*, *Diphylla ecaudata*) and the null allele of *P. mesoamericanus* (electronic supplementary material, figure S5). Under the assumption of a complete removal of functional constraints at  $t$  million years ago (Ma), we applied two methods to estimate the dates of pseudogenization events [34,35]. In the first method, we estimated a posterior probability distribution of  $t$  based on changes in  $\omega$  (electronic supplementary material, figure S5) [34]. In the second method, we counted the existing ORF-disrupting mutations and calculated the waiting times for generating current number ( $n$ ) and future number ( $n + 1$ ) of ORF-disrupting mutations (see also electronic supplementary material). Another posterior probability distribution of  $t$  based on the waiting times was estimated (electronic supplementary material, figure S5). Afterwards, we combined

the two distributions and obtained a final posterior probability distribution of  $t$  (electronic supplementary material, figure S5). Based on these methods, we inferred the dates of *SWS1* pseudogenizations in the following lineages as follows: *Desmodus rotundus* (mode: 4.7 Ma, mean: 5.5 Ma, median: 5.2 Ma, 95% confidence interval: 2.2–10.1 Ma), *Diaemus youngi* (2.6 Ma, 3.6 Ma, 3.1 Ma, 0.8–8.0 Ma), *Diphylla ecaudata* (2.5 Ma, 3.9 Ma, 3.5 Ma, 0.9–8.4 Ma) and *P. mesoamericanus* (1.0 Ma, 2.3 Ma, 1.9 Ma, 0.2–5.9 Ma) (electronic supplementary material, figure S5).

## 4. Discussion

Sensory trade-offs involve specialization in one sensory modality that may lead to the reduction or absence of other sensory modalities. An earlier study revealed a sensory trade-off between an origin of the HDC echolocation and a reduction of colour vision in the Rhinolophidae + Hipposideridae lineage from one of the two suborders of bats, Yinpterochiroptera [5]. Through identifying *SWS1* pseudogenes, the present study revealed the same sensory trade-off in the independently evolved HDC echolocator from the other suborder of bats (Yangochiroptera). In Yangochiroptera, we additionally found a novel sensory trade-off between the gain of the infrared sense and the reduction of colour vision in all three vampire bats.

In contrast to most New World bats (figure 1), the New World HDC bat (*Pteronotus mesoamericanus*) was found to have a pseudogenized allele of *SWS1* in four individuals, while one individual has both pseudogenized alleles and the sixth individual carries both intact alleles (figure 2; electronic supplementary material, figure S4), suggestive of an ongoing pseudogenization. It is unknown whether the intact allele of *SWS1* in the four individuals of *P. mesoamericanus* is functional, but the functional constraint of *SWS1* on this species appears to have relaxed. Interestingly, the allelic polymorphism in the *SWS1* (one allele is pseudogenized, while the other allele is intact) has not been reported in animals previously. This novel polymorphism prompts us to suggest that multiple individuals of a species should be included in examinations of animal opsin genes. Furthermore, we did not identify any ORF-disrupting mutations in the congeneric species *P. davyi* that uses the LDC echolocation (figure 2; electronic supplementary material, figure S4), suggesting relaxation of *SWS1* should have occurred specifically in the HDC echolocator (*P. mesoamericanus*) (electronic supplementary material, figure S4). Despite this, we are not able to rule out the possibility that the null allele may have occurred in the common ancestor of subgenus *Phyllodia* within *Pteronotus*, which included all species previously recognized as the single species, *P. parnelli* [21]. One reason is that due to the absence of samples of species belonging to the subgenus *Phyllodia* within *Pteronotus* such as *P. parnelli* and *P. rubiginosus*, these species may possess shared mutations with *P. mesoamericanus*. Another reason is that the common ancestor of subgenus *Phyllodia* diverged very recently [18], the relaxation of functional constraint may have not had enough time to spread the ORF-disruptive mutation in populations. Certainly, it is possible that changes in *SWS1* transcription or translation are shared among all species within subgenus *Phyllodia*, which may have caused loss of function in their common ancestor. The independent origin of the novel sensory modality (HDC

echolocation) in the New World bats is broadly coincident with the relaxation of *SWS1* evolution within a same time-frame, which parallels with the sensory trade-off between the loss of *SWS1* (i.e. loss of colour vision) and an origin of the HDC echolocation in the Old World bats [5]. That the same sensory trade-off is replicated in two different lineages of HDC echolocating bats (figure 1) that evolved this highly sophisticated form of echolocation independently is indicative of convergent evolution resulting from the same selective pressure: HDC echolocation.

The *SWS1* opsin gene was found to be pseudogenized in all three vampire bats (figure 2; electronic supplementary material, figure S4), suggesting that they lack colour vision. The absence of common ORF-disrupting mutations in vampire bats and purifying selection on the most recent common ancestor of vampire bats suggest that pseudogenization of *SWS1* has occurred recently and independently in the three vampire bat lineages (*Desmodus*, *Diaemus* and *Diphylla*); the ancestral Desmodontinae bats may have retained a functional *SWS1*. Indeed, this inference was supported by our molecular dating of pseudogenization events (electronic supplementary material, figure S5). However, it remains possible that changes in *SWS1* transcription or translation are shared among the three vampire bats, which may have led to functional loss of *SWS1* in their common ancestor. Why has *SWS1* been lost in all three vampire bats? These bats are well known for their blood-feeding habits, but they are also noteworthy because of the presence of infrared sensors [37]. Vampire bats are the only mammals known to use an infrared sense, which they use to detect the warm skin temperatures of endothermic prey and locate the optimal bite sites [11]. The function of this sensory modality may have rendered the dichromatic vision unimportant, which may have led to relaxation of functional constraints on *SWS1*. As such, we propose a sensory trade-off between a gain of the heat sense and a loss of colour vision in vampire bats. Certainly, the gain of the heat sense may also lead to losses of other senses such as the sense of taste [36,38,39], possibly due to the huge energy investment devoted to maintenance and use of a particular sense [40].

Combined with results of earlier studies, our data provide a broader picture of the evolution of vision in bats that are considered sensory specialists. Although dichromatic colour vision is widespread among bats, independent losses of colour vision have now been identified in several lineages (figure 1). These lineages seem to fall into three broad groups. The first group includes the cave-dwelling Old World fruit bats (Pteropodidae), which roost during the day in dark caves in stark contrast with their tree-roosting relatives that retain dichromatic vision (figure 1) [5]. The second group includes two divergent lineages that employ HDC echolocation: an Old World lineage (Hipposideridae + Rhinolophidae), and a New World lineage (*P. mesoamericanus* within Mormoopidae) (figure 1). A third group comprises the three extant vampire bats (subfamily Desmodontinae within Phyllostomidae) (figure 1). These findings highlight that different taxon-specific ecological reasons may lead to the loss of colour vision in different bat lineages [10]. While each of these observations represents only a correlation between a gain of one sense and a loss of another, rather than direct evidence of causation, together they provide a compelling picture supportive of the sensory trade-off hypothesis due to replicated patterns of evolution of sensory modalities in phylogenetically divergent lineages. Together with previous

analyses, our study suggests that evolutionary shifts in animal sensory systems are more frequent than previously thought.

It should be noted that the loss of opsin gene function may have occurred at different stages of protein synthesis: pseudogenization of coding sequences, transcriptional or translational changes. Due to preservation methods, our bat samples from the American Museum of Natural History were only suitable for DNA sequencing, so we were not able to measure mRNA and protein abundances. However, our inference from genetic data was confirmed by mRNA and protein expression data generated by RNA sequencing and immunohistochemical staining [41]. In particular, the SWS1 protein was not detected in the New World high-duty-cycle echolocating bat species (*Pteronotus parnellii*) [41], which belongs to the same species complex as *Pteronotus mesoamericanus* examined in our study [20], suggesting a loss of SWS1 function. Both the SWS1 mRNA transcript and protein were not detected in the common vampire bat (*Desmodus rotundus*), and the SWS1 protein was not found in the other vampire bat (*Diaemus youngi*) [41], suggesting that both vampire bat species have lost SWS1 function. Despite the consistency between DNA and protein, discordance between DNA and protein was also observed in two species (*Trachops cirrhosus* and *Pteronotus davyi*) in which SWS1 protein was not detected but the gene

is intact [41]. The discordance suggests that in some cases the loss of SWS1 function was not caused by changes in its coding sequence, and that changes in gene transcription or translation may have preceded changes in coding sequence and also resulted in its functional loss [41]. Thus, we cannot preclude the possibility that SWS1 function may have been lost in other bat species with an intact coding sequence, and call for further investigations involving examination of mRNA and protein levels to address this possibility in the future.

**Data accessibility.** DNA sequences: GenBank accessions MG251342–MG251380.

**Authors' contributions.** H.Z. and Q.L. designed research; H.J. and J.W. performed research; J.W., H.J., Q.L. and H.Z. analysed data; N.B.S. provided bat tissues and revised the manuscript; and J.W., Q.L. and H.Z. wrote the manuscript.

**Competing interests.** The authors declare that they have no competing interests.

**Funding.** This work was supported in part by the National Natural Science Foundation of China (91331115 and 31722051).

**Acknowledgements.** We are grateful to J. Zhang (University of Michigan) for support in loaning the bat tissues. We thank S. Wang (Wuhan University) for technical assistance in the laboratory.

## References

1. Au WWL, Simmons JA. 2007 Echolocation in dolphins and bats. *Phys. Today* **60**, 40.
2. Kurten L, Schmidt U. 1982 Thermoperception in the common vampire bat (*Desmodus rotundus*). *J. Comp. Physiol. A* **146**, 223–228. (doi:10.1007/BF00610241)
3. Feng P, Zheng J, Rossiter SJ, Wang D, Zhao H. 2014 Massive losses of taste receptor genes in toothed and baleen whales. *Genome Biol. Evol.* **6**, 1254–1265. (doi:10.1093/gbe/evu095)
4. Jones G, Teeling EC, Rossiter SJ. 2013 From the ultrasonic to the infrared: molecular evolution and the sensory biology of bats. *Front. Physiol.* **4**, 117. (doi:10.3389/fphys.2013.00117)
5. Zhao H, Rossiter SJ, Teeling EC, Li CJ, Cotton JA, Zhang S. 2009 The evolution of color vision in nocturnal mammals. *Proc. Natl. Acad. Sci. USA* **106**, 8980–8985. (doi:10.1073/pnas.0813201106)
6. Simmons NB. 2005 *Mammal species of the world: a taxonomic and geographic reference*. Baltimore, MD: Johns Hopkins University Press.
7. Jones G, Teeling EC. 2006 The evolution of echolocation in bats. *Trends Ecol. Evol.* **21**, 149–156. (doi:10.1016/j.tree.2006.01.001)
8. Arita HT, Fenton MB. 1997 Flight and echolocation in the ecology and evolution of bats. *Trends Ecol. Evol.* **12**, 53–58. (doi:10.1016/S0169-5347(96)10058-6)
9. Simmons NB, Geisler JH. 1998 Phylogenetic relationships of *Icaronycteris*, *Archaeonycteris*, *Hassianycteris*, and *Palaeochiropteryx* to extant bat lineages, with comments on the evolution of echolocation and foraging strategies in Microchiroptera. *Bull. Am. Mus. Nat. Hist.* **235**, 1–182.
10. Jacobs GH. 2013 Losses of functional opsin genes, short-wavelength cone photopigments, and color vision—a significant trend in the evolution of mammalian vision. *Vis. Neurosci.* **30**, 39–53. (doi:10.1017/S0952523812000429)
11. Kunz TH, Fenton MB. 2003 *Bat ecology*. Chicago, IL: University of Chicago Press.
12. Peichl L. 2005 Diversity of mammalian photoreceptor properties: adaptations to habitat and lifestyle? *Anat. Rec.* **287A**, 1001–1012. (doi:10.1002/ar.a.20262)
13. Cowing JA, Arrese CA, Davies WL, Beazley LD, Hunt DM. 2008 Cone visual pigments in two marsupial species: the fat-tailed dunnart (*Sminthopsis crassicaudata*) and the honey possum (*Tarsipes rostratus*). *Proc. R. Soc. B* **275**, 1491–1499. (doi:10.1098/rspb.2008.0248)
14. Davies WL, Carvalho LS, Cowing JA, Beazley LD, Hunt DM, Arrese CA. 2007 Visual pigments of the platypus: a novel route to mammalian colour vision. *Curr. Biol.* **17**, R161–R163. (doi:10.1016/j.cub.2007.01.037)
15. David-Gray ZK, Bellingham J, Munoz M, Avivi A, Nevo E, Foster RG. 2002 Adaptive loss of ultraviolet-sensitive/violet-sensitive (UVS/VS) cone opsin in the blind mole rat (*Spalax ehrenbergi*). *Eur. J. Neurosci.* **16**, 1186–1194. (doi:10.1046/j.1460-9568.2002.02161.x)
16. Tan Y, Yoder AD, Yamashita N, Li W-H. 2005 Evidence from opsin genes rejects nocturnality in ancestral primates. *Proc. Natl. Acad. Sci. USA* **102**, 14 712–14 716. (doi:10.1073/pnas.0507042102)
17. Datzmann T, von Helversen O, Mayer F. 2010 Evolution of nectarivory in phyllostomid bats (Phyllostomidae Gray, 1825, Chiroptera: Mammalia). *BMC Evol. Biol.* **10**, 165. (doi:10.1186/1471-2148-10-165)
18. Rojas D, Warsi OM, Dávalos LM. 2016 Bats (Chiroptera: Noctilionoidea) challenge a recent origin of extant neotropical diversity. *Syst. Biol.* **65**, 432–448. (doi:10.1093/sysbio/syw011)
19. Dávalos LM, Cirranello AL, Geisler JH, Simmons NB. 2012 Understanding phylogenetic incongruence: lessons from phyllostomid bats. *Biol. Rev.* **87**, 991–1024. (doi:10.1111/j.1469-185x.2012.00240.x)
20. Clare EL, Adams AM, Maya-Simões AZ, Eger JL, Hebert PD, Fenton MB. 2013 Diversification and reproductive isolation: cryptic species in the only New World high-duty cycle bat, *Pteronotus parnellii*. *BMC Evol. Biol.* **13**, 26. (doi:10.1186/1471-2148-13-26)
21. Pavan AC, Marroig G. 2016 Integrating multiple evidences in taxonomy: species diversity and phylogeny of mustached bats (Mormoopidae: *Pteronotus*). *Mol. Phylogenet. Evol.* **103**, 184–198. (doi:10.1016/j.ympev.2016.07.011)
22. Hall TA. 1999 BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp. Ser.* **41**, 95–98.
23. Huelsenbeck J, Ronquist F. 2001 MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* **17**, 754–755. (doi:10.1093/bioinformatics/17.8.754)
24. Posada D, Crandall K. 1998 MODELTEST: testing the model of DNA substitution. *Bioinformatics* **14**, 817–818. (doi:10.1093/bioinformatics/14.9.817)
25. Posada D. 2008 jModelTest: phylogenetic model averaging. *Mol. Biol. Evol.* **25**, 1253–1256. (doi:10.1093/molbev/msn083)

26. Schmidt HA, Strimmer K, Vingron M, von Haeseler A. 2002 TREE-PUZZLE: maximum likelihood phylogenetic analysis using quartets and parallel computing. *Bioinformatics* **18**, 502–504. (doi:10.1093/bioinformatics/18.3.502)

27. Nei M, Gojobori T. 1986 Simple methods for estimating the numbers of synonymous and nonsynonymous nucleotide substitutions. *Mol. Biol. Evol.* **3**, 418–426. (doi:10.1093/oxfordjournals.molbev.a040410)

28. Pride D. 2004 SWAAP 1.0.2: a tool for analyzing substitutions and similarity in multiple alignments. See <http://www.bacteriamuseum.org/SWAAP/SwaapPage.htm>

29. Yang Z. 2007 PAML 4: phylogenetic analysis by maximum likelihood. *Mol. Biol. Evol.* **24**, 1586–1591. (doi:10.1093/molbev/msm088)

30. Yang Z, Kumar S, Nei M. 1995 A new method of inference of ancestral nucleotide and amino acid sequences. *Genetics* **141**, 1641–1650.

31. Zhang J, Nei M. 1997 Accuracies of ancestral amino acid sequences inferred by the parsimony, likelihood, and distance methods. *J. Mol. Evol.* **44**, S139–S146. (doi:10.1007/Pl00000067)

32. Wertheim JO, Murrell B, Smith MD, Pond SLK, Scheffler K. 2015 RELAX: detecting relaxed selection in a phylogenetic framework. *Mol. Biol. Evol.* **32**, 820–832. (doi:10.1093/molbev/msu400)

33. Pond SLK, Frost SDW, Muse SV. 2005 HyPhy: hypothesis testing using phylogenies. *Bioinformatics* **21**, 676–679. (doi:10.1093/bioinformatics/bti079)

34. Meredith RW, Gatesy J, Murphy WJ, Ryder OA, Springer MS. 2009 Molecular decay of the tooth gene *enamelin (ENAM)* mirrors the loss of enamel in the fossil record of placental mammals. *PLoS Genet.* **5**, e1000634. (doi:10.1371/journal.pgen.1000634)

35. Zhao H, Yang J-R, Xu H, Zhang J. 2010 Pseudogenization of the umami taste receptor gene *Tas1r1* in the giant panda coincided with its dietary switch to bamboo. *Mol. Biol. Evol.* **27**, 2669–2673. (doi:10.1093/molbev/msq153)

36. Zhao H, Zhou Y, Pinto CM, Charles-Dominique P, Galindo-Gonzalez J, Zhang S, Zhang J. 2010 Evolution of the sweet taste receptor gene *Tas1r2* in bats. *Mol. Biol. Evol.* **27**, 2642–2650. (doi:10.1093/molbev/msq152)

37. Gracheva EO, Cordero-Morales JF, González-Carcácia JA, Ingolia NT, Manno C, Aranguren CI, Weissman JS, Julius D. 2011 Ganglion-specific splicing of *TRPV1* underlies infrared sensation in vampire bats. *Nature* **476**, 88–91. (doi:10.1038/nature10245)

38. Zhao H, Xu D, Zhang S, Zhang J. 2012 Genomic and genetic evidence for the loss of umami taste in bats. *Genome Biol. Evol.* **4**, 73–79. (doi:10.1093/gbe/evr126)

39. Hong W, Zhao H. 2014 Vampire bats exhibit evolutionary reduction of bitter taste receptor genes common to other bats. *Proc. R. Soc. B* **281**, 20171079. (doi:10.1098/rspb.2014.1079)

40. Niven JE, Laughlin SB. 2008 Energy limitation as a selective pressure on the evolution of sensory systems. *J. Exp. Biol.* **211**, 1792–1804. (doi:10.1242/jeb.017574)

41. Sadier A *et al.* 2018 Evidence for multifactorial processes underlying phenotypic variation in bat visual opsins. *bioRxiv*. (doi:10.1101/300301)